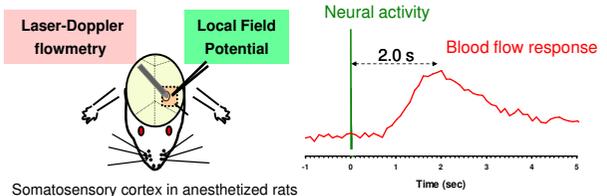
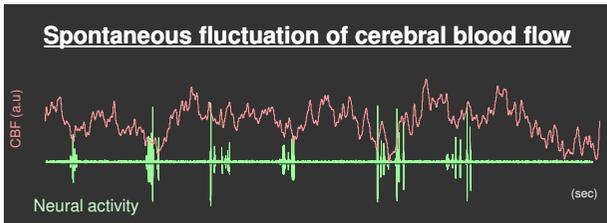


### 脳機能イメージングと脳微小循環計測

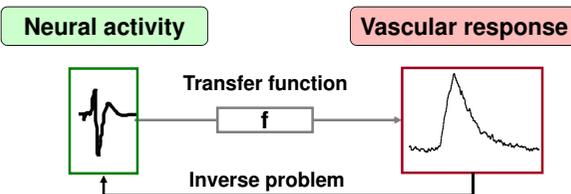


正本 和人<sup>1,2,3</sup>  
<sup>1</sup>電気通信大学 先端領域教育研究センター 特任准教授  
<sup>2</sup>放射線医学総合研究所 分子イメージング研究センター 客員協力研究員  
<sup>3</sup>慶應義塾大学 医学部 神経内科 訪問助教

UEC Tokyo  
 (The University of Electro-Communications)



### Physiological coupling of Neural and Vascular activities

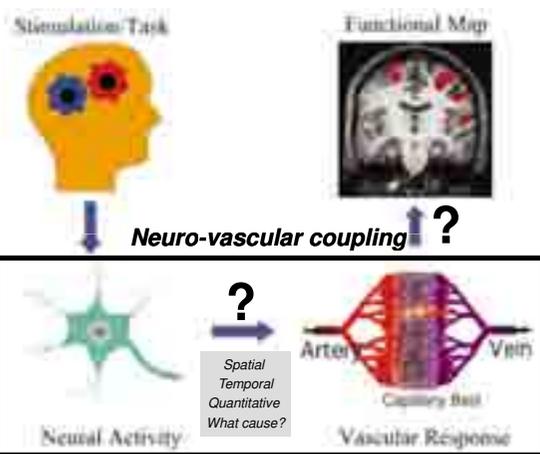
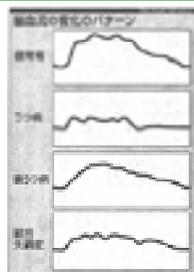


We can see brain functions noninvasively by measuring vascular responses (not electrical activity!)

### Optical imaging of human brain function



(Hasegawa and Sato, 2007)



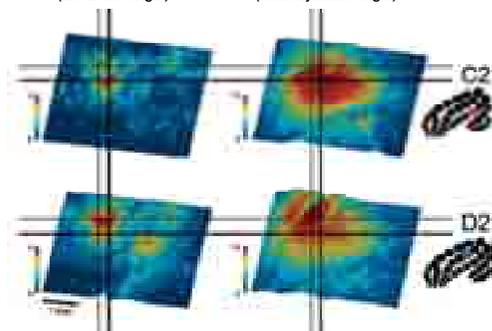
### Understanding of Neuro-Vascular interactions



- 1. Spatiotemporal accuracy (時間空間の特異性)**  
How accurate blood vessels respond to neural activity?
- 2. Quantitative relationship (定量性)**  
Larger vascular response means stronger neural activity?
- 3. Physiologic mechanism (生理学的機構)**  
How can we interpret the vascular signals in terms of neural functions?

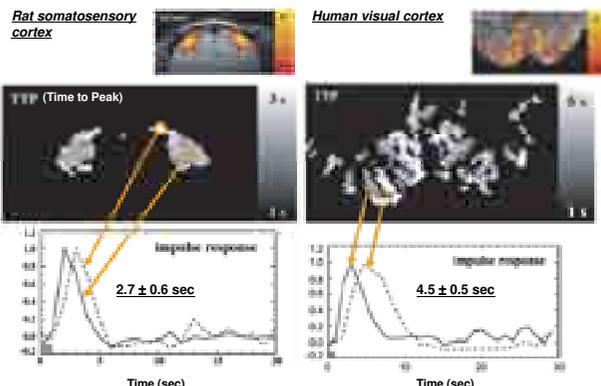
### Spatial specificity of metabolic and hemodynamic signals

Auto fluorescence image (Metabolic origin)      Laser speckle imaging (Hemodynamic origin)

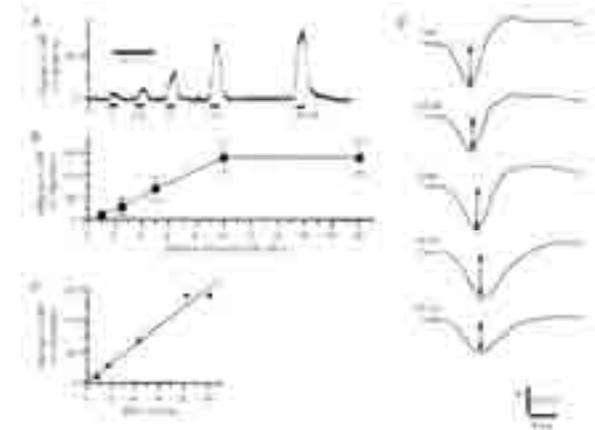


(Webner et al., 2004)

### Species differences in hemodynamic response function

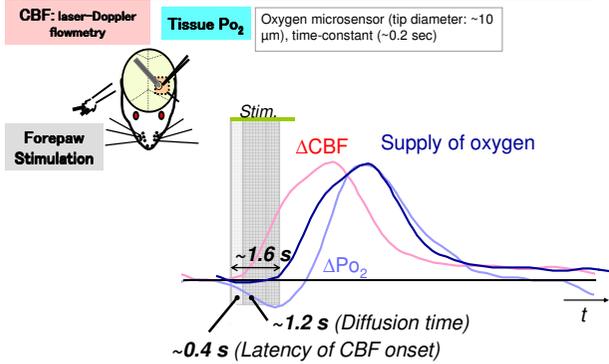


(de Zwart et al., 2005)



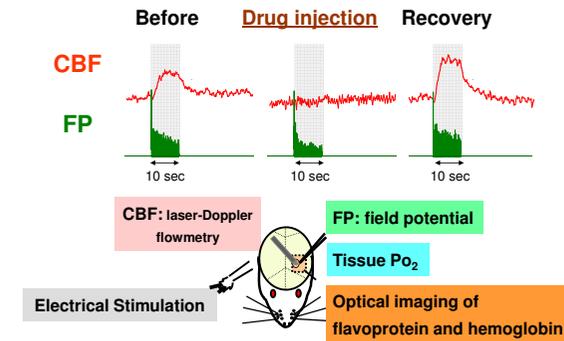
(Mathiesen et al., 1998)

### Lags behind Demand due to Diffusion Time for Oxygen



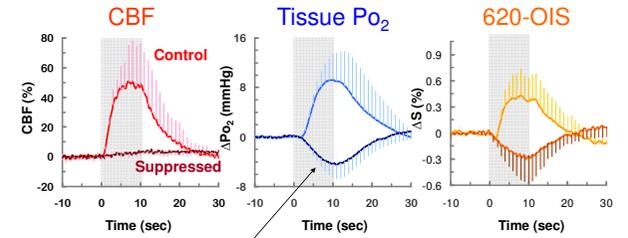
(Masamoto et al., JAP 2007)

### Suppressed vascular response experiment



(Masamoto et al., 2007)

### Decrease in oxygen tension due to CMRO2 increase



Tissue and blood  $P_{O_2}$  decrease due to an increase in oxygen consumption (CMRO2)

(Masamoto et al., NeuroImage 2007)

### Dynamic CMRO2 with Oxygen Exchange Model

$$V_c \frac{dC_c(t)}{dt} = \underbrace{2CBF(t)(C_a - C_c(t))}_{\text{Supply}} - \underbrace{PS(C_p(t) - C_c(t))}_{\text{Exchange}}$$

$$V_t \frac{dC_t(t)}{dt} = \underbrace{PS(C_p(t) - C_t(t))}_{\text{Exchange}} - \underbrace{CMRO_2(t)}_{\text{Demand}}$$

$$C_c(t) = C_p(t) + \frac{4[Hb]}{1 + \left(\frac{P_{50}}{C_c(t)}\right)^{hill}}$$

Capillary

Tissue

Plasma ( $C_p$ ) and hemoglobin-binding oxygen

CMRO<sub>2</sub>

Tissue  $P_{O_2}$

Constant:

$V_c = 1$  mL/100 g (capillary)  
 $V_t = 97$  mL/100 g (tissue)  
 $\alpha = 0.00139$  mmol/L/mmHg  
 $P_{50} = 38$  mmHg, hill = 2.73  
 $PS = 7000$  mL/min/100g  
 $CBF = 150$  mL/min/100 g\*  
 (\*ASL- MRI technique)

Measured:

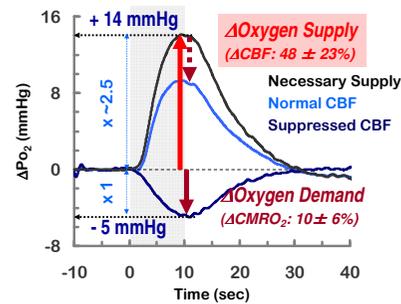
CBF (t): LDF (t) x CBF  
 $C_t$  (t): tissue  $P_{O_2}$   
 $C_a$ , [Hb]: systemic arterial blood

Variable:

CMRO<sub>2</sub> (t) (mL/min/100 g)  
 $C_c$  (t): capillary oxygen concentration

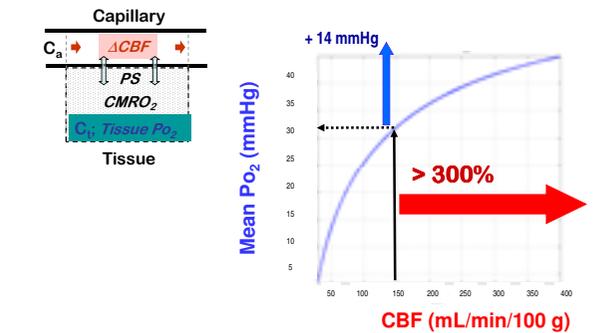
(Modified from Valabregue et al., JCBFM 2003)

### 2.5 Times Higher Supply Relative to Demand

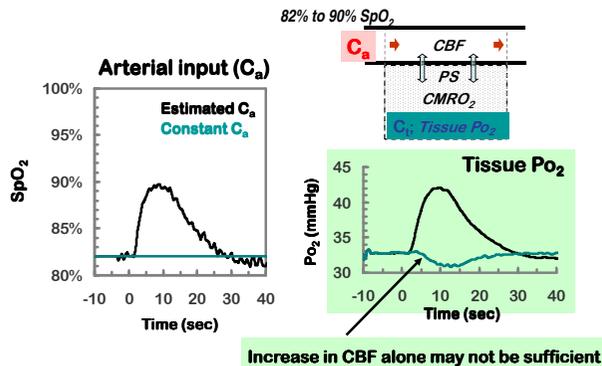


(Masamoto et al., 2007)

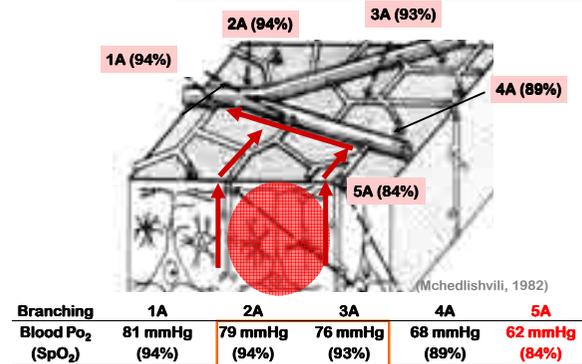
### Po2 increase is not explainable by the change in CBF alone



### Dynamic changes in arterial oxygenation level

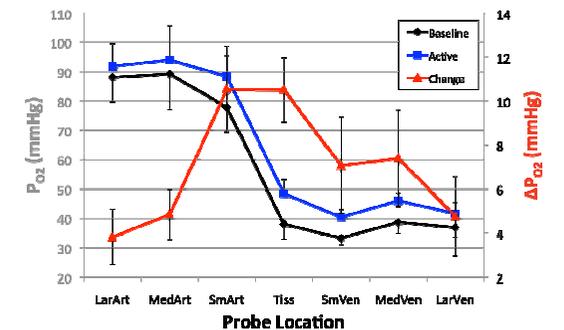


### Recruitment of upstream arterial blood

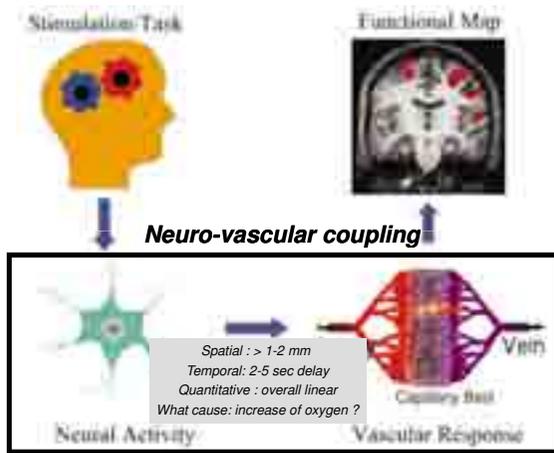


( $P_{O_2}$  and SpO<sub>2</sub> data from Vovenko, 1999)

### PO2 Changes in vascular and tissue compartment



(Vazquez et al., JCBFM 2010)



### テニユア・トラックにおける研究提案

(検討項目)

1. 血流反応の時間空間的特異性
2. 神経-血流反応の定量性
3. 血管反応のメカニズム (神経科学的解釈)

(具体的アプローチ)

#### ① 動物実験モデルの構築

長期に渡り継続観察可能なマウス・ラットを用いた動物実験モデルの構築

#### ② 可視化手法の確立

脳血管形態を可視化するための光イメージング手法の確立

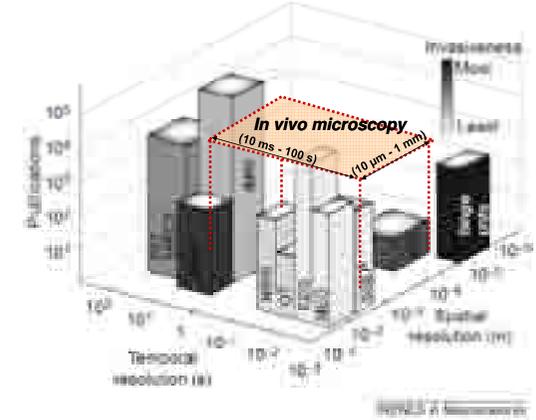
脳内環境 (酸素・グルコース) の光イメージング手法の確立

#### ③ 生体活動の計測

脳神経活動に対する血管反応の領域・大きさに関する検討

様々な神経活動に対するエネルギー代謝・血管反応の計測

### Comparison of functional brain imaging tools



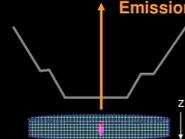
### Two-Photon Laser Scanning Fluorescence Microscopy

1. Deep penetration
2. Less photobleaching and photodamages

Near infrared laser (700-1000 nm wavelength)



Detection with PMT

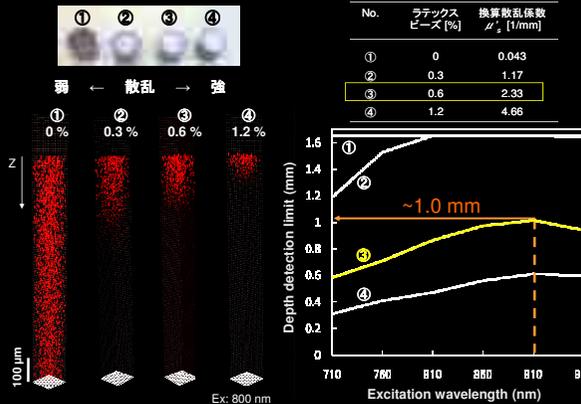


(Raster scanning)

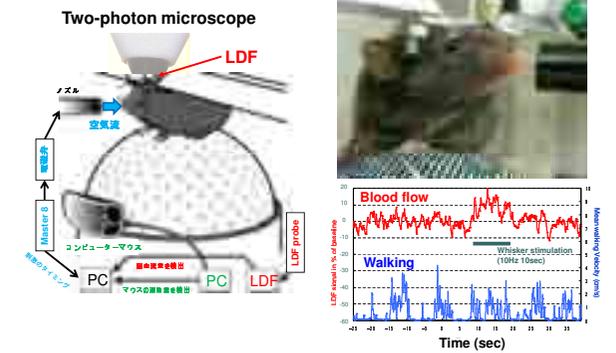
16,000 lines/sec  
30 fps (512 x 512)  
60 fps (512 x 256)  
100 fps (512 x 128)  
200 fps (512 x 64)

Agar (2.0%) with fluorescent micro-beads (1  $\mu\text{m}$ )

### Tissue Scattering Effect on Depth Detection Limit



### Dynamic CBF measurements in awake mice



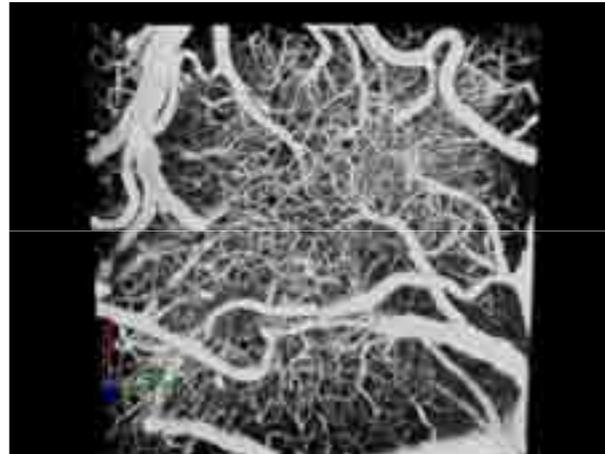
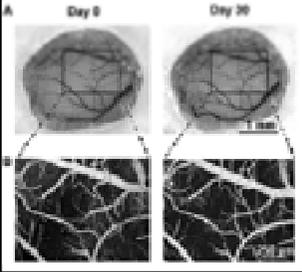
(Takuwa et al., submitted)

### Two-photon imaging through a closed cranial window

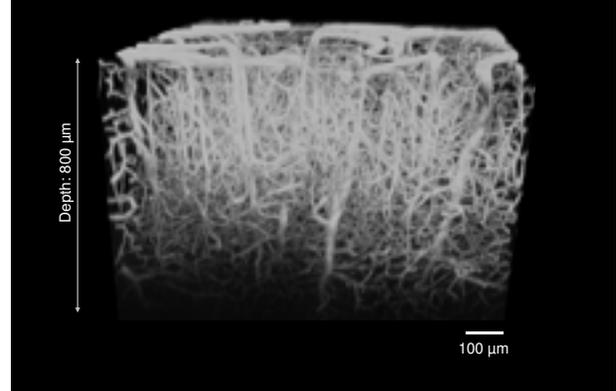
C57BL/6J mice (20-30 g)  
Isoflurane (1%) or awake  
Closed cranial window: open skull (3 mm in diameter), dura intact  
Fluorescent dye: Sulforhodamine 101 (5-10 mM in saline, 8 ml/kg)

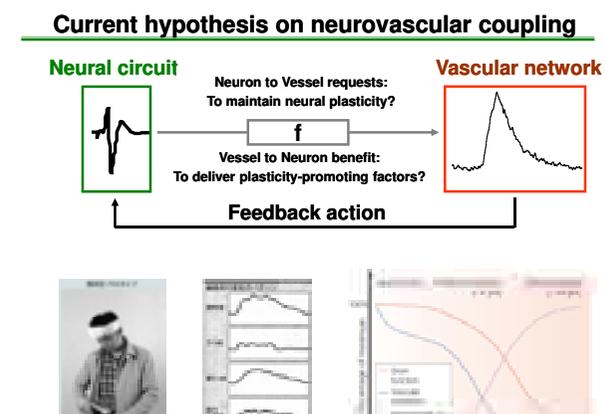
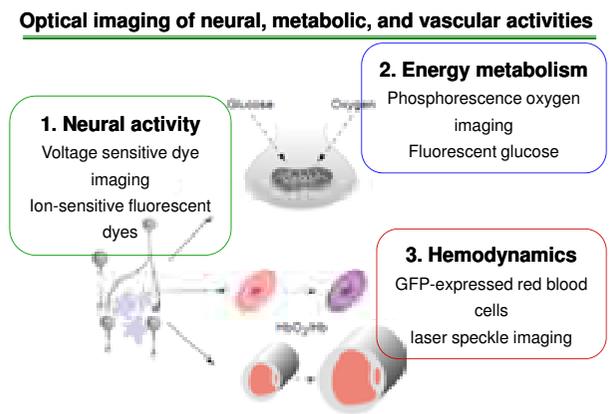
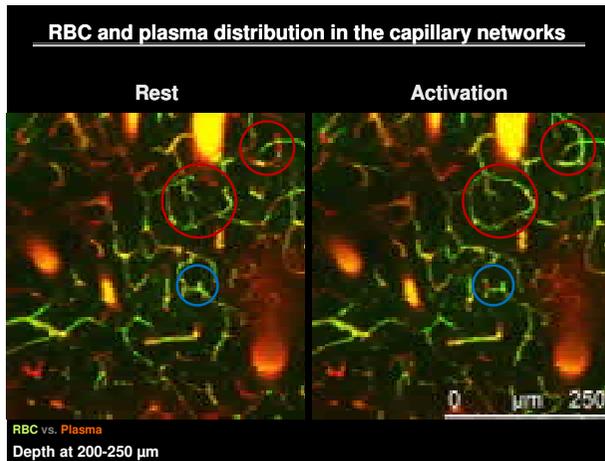
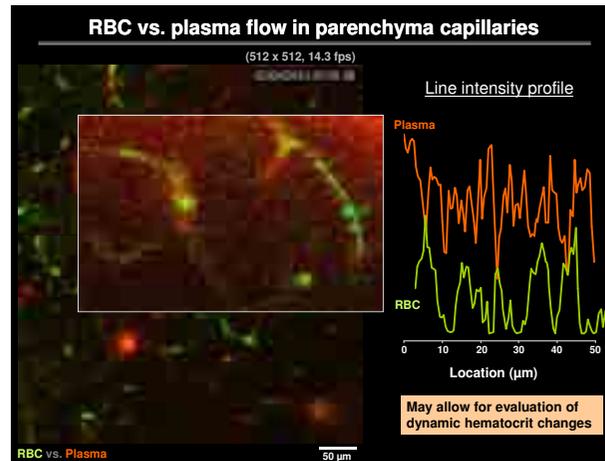
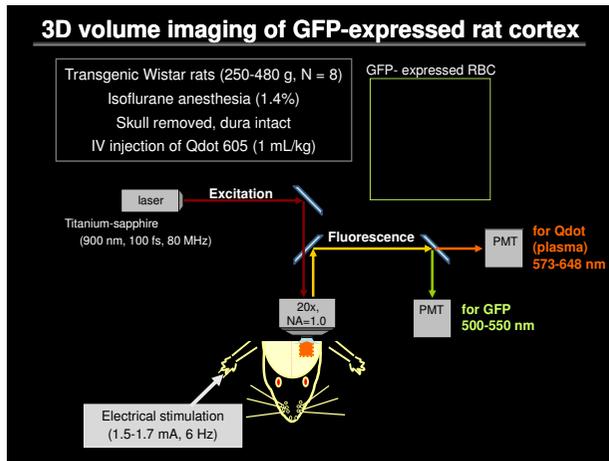
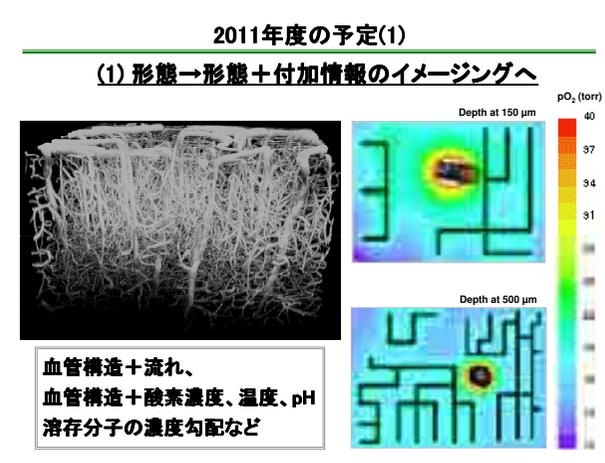
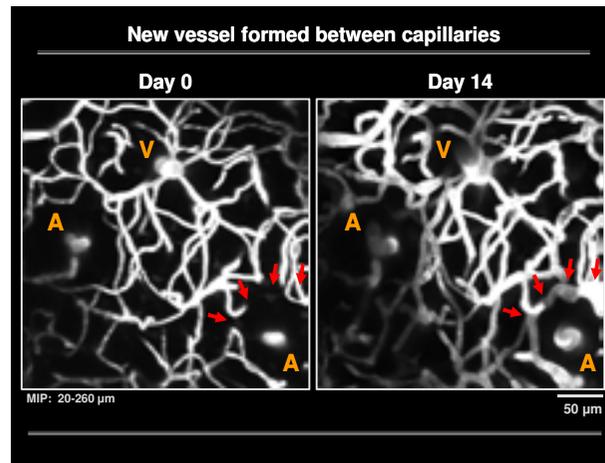
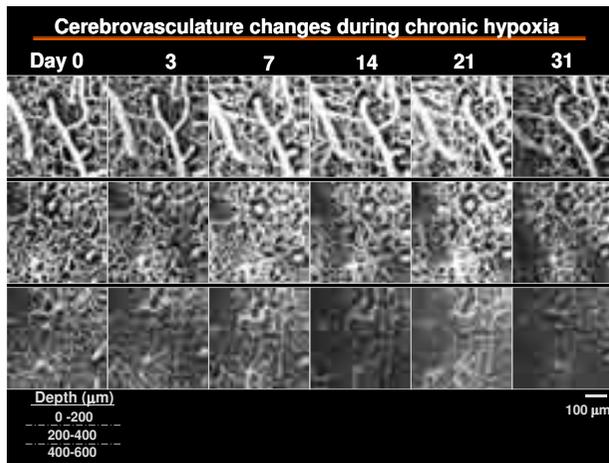


Y. Tomita et al., JCBFM (2005) 858-867  
Long-term in vivo visualization of neural microcirculation in the cerebral cortex of awake mice by the use of neural endoscopes



### Imaging of vascular networks in live mice cerebral cortex





### 謝辞

(敬称略)

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